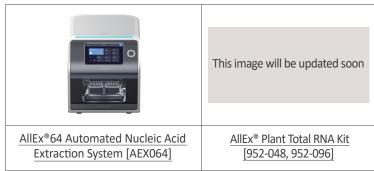
Proposal of optimal sample amounts for high-quality total RNA extraction from sweet potato using the AllEx® Mini Plant Total RNA Kit and AllEx® Plant Total RNA Kit

Experimental Conditions

Materials





Sample & Extraction Information

| Origin | Ipomoea trifida L. | | |
|----------------------------|---|---------|--|
| Sample | | | |
| | Sweet potato (<i>Ipomoea batatas</i> L.) | | |
| Target | Total RNA | | |
| Sample amount | 100, 150, 200 mg | | |
| Elution volume | Up to 100 μl | | |
| Extraction system protocol | Plant RNA | | |
| Operating time | AllEx® Mini | 23' 04" | |
| Operating time | AllEx®64 | 22′ 30″ | |

Protocol

AllEx® Mini Plant Total RNA Kit & AllEx® Plant Total RNA Kit Protocol

* For more details and methods, please refer to the manual of AllEx® Mini Plant Total RNA Kit or AllEx® Plant Total RNA Kit.

Preparation of DNase I Solution

To obtain a DNase I solution of 2 Kunitz units/ μ I, add 120 μ I of Nuclease-free water to the tube containing lyophilized DNase I (240 Kunitz units), and mix carefully and gently to avoid foaming. The reconstituted enzyme should be stored at -20 °C for long-term stability.

Sample Preparation

- 1. Place 100, 150, or 200 mg of the samples into a 2 ml microcentrifuge tube containing a 5 mm stainless steel bead.
- 2. Grind each sample into a fine powder using a TissueLyser II (or an equivalent bead-beater) at 30 Hz for 1 min.
- 3. Add Buffer SQ1 to each sample in the follows:

| Sample amount | Buffer SQ1 | | |
|---------------|------------|--|--|
| 100 mg | 400 μΙ | | |
| 150 mg | 500 μΙ | | |
| 200 mg | 600 นไ | | |

- 4. Vortex vigorously for 30 s and incubate for 5 min at room temperature.
- 5. Vortex vigorously and centrifuge at 13,000 rpm for 10 min at 4 °C.
- 6. Carefully transfer 300 μl of the supernatant to the 1st (7th) well of the cartridge, avoiding the starch and debris layer.
- 7. Add 4 μ l of DNase I solution (2 Kunitz units/ μ l) to the 3rd (9th) well of the cartridge.

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Results

| | Sweet potato (Ipomoea batatas L.) | Mean (n=2) | | | | | |
|---|--------------------------------------|---------------------------------|-------------|----------------------------|-------------|-------------|-------------|
| | | AllEx® Mini Plant Total RNA Kit | | AllEx® Plant Total RNA Kit | | | |
| | | 100 mg (CV) | 150 mg (CV) | 200 mg (CV) | 100 mg (CV) | 150 mg (CV) | 200 mg (CV) |
| П | Yield (µg) | 29.4 (0.14) | 35.8 (0.03) | 33.5 (0.01) | 36.7 (0.25) | 37.9 (0.06) | 31.3 (0.13) |
| | A ₂₆₀ /A ₂₈₀ | 2.19 (0.00) | 2.19 (0.00) | 2.19 (0.00) | 2.19 (0.00) | 2.20 (0.00) | 2.20 (0.00) |
| | A ₂₆₀ /A ₂₃₀ | 2.31 (0.01) | 2.32 (0.00) | 2.32 (0.01) | 2.33 (0.01) | 2.35 (0.01) | 2.32 (0.01) |

Table 1. Evaluation of RNA yield and purity from sweet potato

Total RNA was extracted from 100, 150, or 200 mg of sweet potato (n=2) using the AllEx® Mini Plant Total RNA Kit with the AllEx® Mini Automated Nucleic Acid Extraction System or the AllEx® Plant Total RNA Kit with the AllEx®64 Automated Nucleic Acid Extraction System, operating the Plant RNA protocol. The yield and purity of the extracted RNA were evaluated with a NanoDrop™ 2000 spectrophotometer.

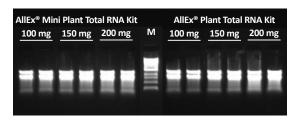


Figure 1. Agarose gel electrophoresis results of extracted total RNA from sweet potato Extracted total RNA samples were subjected to electrophoresis on a 1% agarose gel in 0.5X TBE buffer at 150 V for 15 min. The extracted RNA size was calculated using the GENESTA™ 1 kb DNA Ladder (GA-100), and the electrophoresis results were analyzed with the SmartView Pro 1100 Imager System (UVCI-1100).

Conclusion

- Optimal pretreatment conditions and procedures were established for extracting high-quality total RNA from the starch-enriched tuberous root of sweet potato.
- If the sample is not sufficiently pulverized into powder during the grinding process, the extraction efficiency may be reduced. In such cases, please use liquid nitrogen to completely grind the sample into powder before proceeding to the lysis step.
- Ensuring the appropriate ratio (approximately 1:3) between the sample and Buffer SQ1 is essential for effective extraction.

Ordering Information

| Cat. No. | Product | Size | | |
|----------|---|------|--|--|
| AEX012 | AEX012 AllEx® Mini Automated Nucleic Acid Extraction System | | | |
| 978-048 | 978-048 AllEx® Mini Plant Total RNA Kit (Single Type) | | | |
| AEX064 | AEX064 AllEx®64 Automated Nucleic Acid Extraction System | | | |
| 952-048 | AllEx® Plant Total RNA Kit (Single Type) | 48 T | | |
| 952-096 | AllEx® Plant Total RNA Kit (Plate Type) | 96 T | | |